The Structure of Muscimol, a GABA Analogue of Restricted Conformation

LOTTE BREHM, HANS HJEDS and POVL KROGSGAARD-LARSEN

The Royal Danish School of Pharmacy, Chemical Laboratory C, DK-2100 Copenhagen Ø, Denmark

Intensive studies in recent years have furnished evidence of the role of γ-aminobutyric acid (GABA) as an inhibitory transmitter in the mammalian central nervous system.¹⁻⁴ Some conceptions concerning the structural basis of GABA-receptor interactions have been provided by investigations of the specific GABA antagonist bicucullin.^{2,5} Preliminary X-ray analyses indicate that GABA exists in a partially folded conformation in the solid state,^{5,6} while the results of molecular orbital calculations on GABA,⁷ supported by the GABA-like activity of 4-aminotetrolic acid,⁸ seem to indicate that GABA is in an extended conformation, when it acts on the receptors.

Muscimol, an isoxazole enol-betaine isolated from Amanita muscaria, has a pronounced and multifarious effect, including psychogenic activity, upon the human central nervous system.^{9,10} On certain central interneurons in cats, however, muscimol has been shown to exert a depressant activity very similar to that of GABA.11 Furthermore muscimol, like GABA, is antagonized by bicucullin.2 Thus it is likely to be a GABA analogue of restricted conformation. Correlation of the detailed structures of muscimol and analogues and their corresponding physiological action seems to be a rational approach to elicit the structural characteristics of the GABA receptors.

As part of our investigations an X-ray analysis of muscimol has been performed. Structural determinations of muscimol analogues, and the syntheses of isoxazole cool-betaines with additional restrictions of the conformation are in progress.

Crystals of muscimol, $C_4\hat{H}_6N_2O_2$, were obtained by diffusion of ethanol in aqueous solutions of the compound. By varying the relative concentrations, the compound was obtained in the monoclinic form and the triclinic form. A structure determina-

tion has been made on the monoclinic form, which has space group $P2_1/n$, a=10.738(9), b=6.950(4), c=6.794(7) Å, $\beta=98.18(7)^\circ$, Z=4. Three-dimensional diffraction data in the range $2.5 \le \theta \le 25^\circ$ were measured on a Nonius three-circle automatic diffractometer using graphite monochromated $MoK\alpha$ radiation.

The structure was solved using a modified symbolic addition procedure 12 and refined using the full-matrix least-squares method. The present R value is 0.069

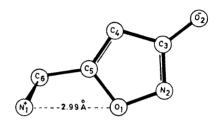


Fig. 1.

The figure illustrates the conformation of the molecules and shows the interatomic distance $N(1)^+ \cdots O(1)$ of 2.99 Å. The muscimol molecule adopts a conformation in which the side chain is directed toward the ring oxygen. This is in accordance with molecular orbital calculations. molecular orbital calculations. On the other hand the side chain is not coplanar with the ring plane, and this observation differs from the molecular orbital predictions. The torsion angle $O(1)-C(5)-C(6)-N(1)^+$ found (60°) is similar to the corresponding torsion angle of muscarine (73°) and to those of other muscarinic agonists.13 The intramolecular $O(2)^- \cdots N(1)^+$ distance is 5.77 Å. The molecular orbital calculations predict $O(2)^-$ and $N(1)^+$ to be 5.0 – 6.0 Å apart in both muscimol and GABA.

The detailed crystal structure of muscimol will be described elsewhere.

Acknowledgements. The authors wish to thank Dr. K. Nakamura, Sankyo Co. Ltd., Tokyo, Japan, for supplying a sample of muscimol, and the Danish Medical Research Council for supporting this work. The authors express their gratitude to Professor Bodil Jerslev for her stimulating interest in this work.

- 1. Krnjević, K. Nature 228 (1970) 119.
- Curtis, D. R., Duggan, A. W., Felix, D. and Johnston, G. A. R. Nature 226 (1970) 1222.
- Curtis, D. R., Duggan, A. W., Felix, D. and Johnston, G. A. R. Nature 228 (1970) 676
- Mitchell, J. F. and Srinivasan, V. Nature 224 (1969) 663.
- Steward, E. G., Player, R., Quilliam, J.P., Brown, D. A. and Pringle, M. J. Nature New Biology 233 (1971) 87.
- 6. Tomita, K. Tetrahedron Letters 1971 2587. 7. Kier, L.B. and Truitt, Jr., E.B. Ex-
- Kier, L. B. and Truitt, Jr., E. B. Ex perientia 26 (1970) 988.
- Beart, P. M., Curtis, D. R. and Johnston,
 G. A. R. Nature New Biology 234 (1971)
 80.
- Waser, P. G. In Efron, D. H., Holmstedt, B. and Kline, N. S. Ethnopharmacologic Search for Psychoactive Drugs, Public Health Service Publication No. 1645, U. S. Government Printing Office, Washington D. C. 1967, p. 419.
- Theobald, W., Büch, O., Kunz, H.A., Krupp, P., Stenger, E. G. and Heimann, H. Arzneimittelforsch. 18 (1968) 311.
- Johnston, G. A. R., Curtis, D. R., DeGroat, W. C. and Duggan, A. W. Biochem. Pharmacol. 17 (1968) 2488.
- Stewart, J. M., Kundell, F. A. and Baldwin, J. C. X-Ray 70 Crystal Structure Calculation System, Computer Science Center, University of Maryland, July 1970.
- University of Maryland, July 1970.

 13. Shefter, E. In Triggle, D. J., Moran, J. F. and Barnard, E. A., Eds., Cholinergic Ligand Interactions, Academic, New York and London 1971, p. 83.

Received March 6, 1972.

Formation of 2-Oxoisovalerate Dehydrogenase in *Pseudomonas* fluorescens

MATTI PUUKKA, PEKKA MÄNTSÄLÄ, HARRI LÖNNBERG, RAIJA PAJULA and VEIKKO NURMIKKO

Department of Biochemistry, University of Turku, Turku, Finland

Decarboxylation of branched chain 2-oxo acids, 2-oxoisovalerate, 2-oxoisocaproate, and 2-oxo-3-methylvalerate has been detected in animal tissues, 1,2 and in Pseudomonas aeruginosa,3 Bacillus subtilis,4 and Pseudomonas putida.5 2-Oxoisovalerate dehydrogenase from Bacillus subtilis has been purified about 40-fold and it catalyses oxidative decarboxylation of all the three branched chain 2-oxo acids, thus yielding the related acyl coenzyme A esters.4

The enzyme is highly stereospecific and the L-isomer is the active substrate. Sulphhydryl reactants inhibit the activity of 2-oxoisovalerate dehydrogenase. The enzyme was induced during growth on valine on Pseudomonas putida.

The present investigation shows that 2-oxoisovalerate dehydrogenase is formed in the presence of valine, isoleucine, leucine, and 2-oxo acids derived from these amino acids in *Pseudomonas fluorescens* (strains P-2 and UK-1).

Materials. I.-Amino acid oxidase and catalase were purchased from Calbiochem, Los Angeles, and 1.¹⁴C-I.-valine from the New England Nuclear Corporation, Boston. 1.¹⁴C-2-Oxoisovalerate was prepared as described earlier by Meister ⁶ and purified by ion exchange chromatography.

Cultures. Pseudomonas fluorescens P-2 and UK-1 were used as test organisms. Ps. fluorescens P-2 was grown with aeration in the salt solution described by Goodhue and Snell with 10 mM of various carbon sources. The strain UK-1 was cultured in the basal salt solution containing 0.817 g of KH₂PO₄, 0.247 g of MgSO₄.7H₂O, and 2.8 mg of FeSO₄.7H₂O per litre.

Growth was estimated from turbidity measurements made with a Klett-Summerson colorimeter, employing filter 62. Cultures were grown as described elsewhere, with some modifications.⁸

Enzyme preparation and assay. The samples (about 4 mg dry weight) withdrawn from the